

Advancing AAV Capsid Engineering for Targeted Gene Therapy Using PackGene's π-Icosa Platform



Introduction

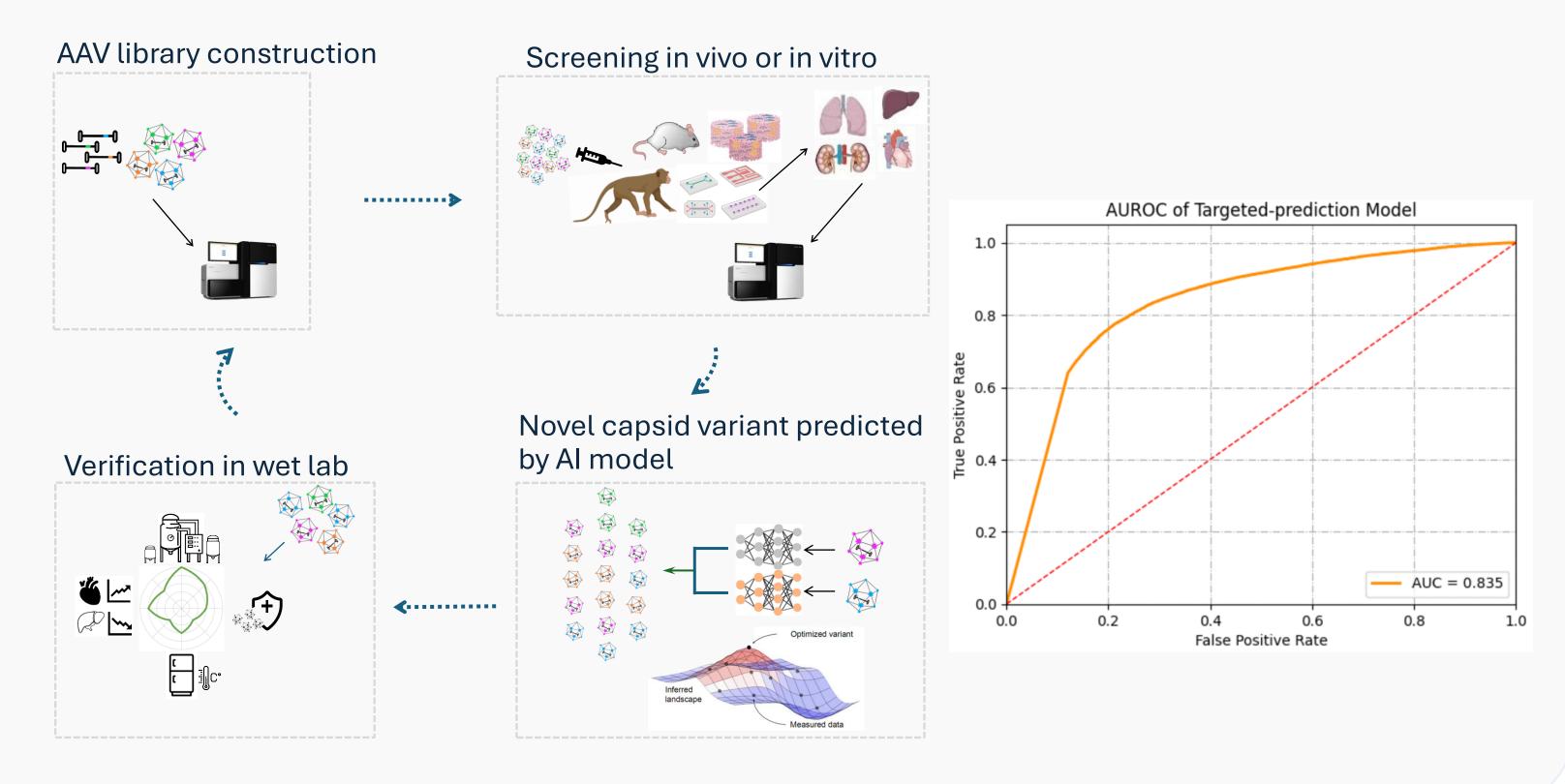
Adeno-associated virus (AAV) vectors are widely utilized in gene therapy for their ability to deliver genetic material to specific tissues with minimal immunogenicity. However, clinical application remains limited by suboptimal tissue targeting, inefficient transduction, and low manufacturability. Traditional methods of capsid development often fall short in addressing these multifaceted challenges. To bridge this gap, PackGene has developed the π -Icosa Capsid Engineering Platform—a next-generation solution that integrates rational design, directed evolution, and artificial intelligence to create capsids with enhanced specificity, efficacy, and production efficiency. This platform is designed to accelerate the discovery of novel AAV variants tailored for therapeutic delivery across CNS, muscle, eye, and immune tissues.

π-Icosa Capsid Engineering Platform

The π -lcosa platform is a multi-phase system for AAV capsid library construction, in vivo screening, and optimization. It enables the identification of top-performing capsid variants with improved tissue tropism and minimized off-target expression.

Key features include:

- Al-Based Capsid Design:
- Incorporates a proprietary transformer-based prediction model pre-trained using protein language models.
- Achieved AUC of 0.83 on directed evolution datasets for tissue-specific targeting.
- Workflow Overview:
 - AAV library construction
 - In vivo/in vitro screening
 - Al-predicted capsid variant selection
 - Experimental validation





Reliable CRO & CDMO for AAV lentivirus · mRNA · plasmid

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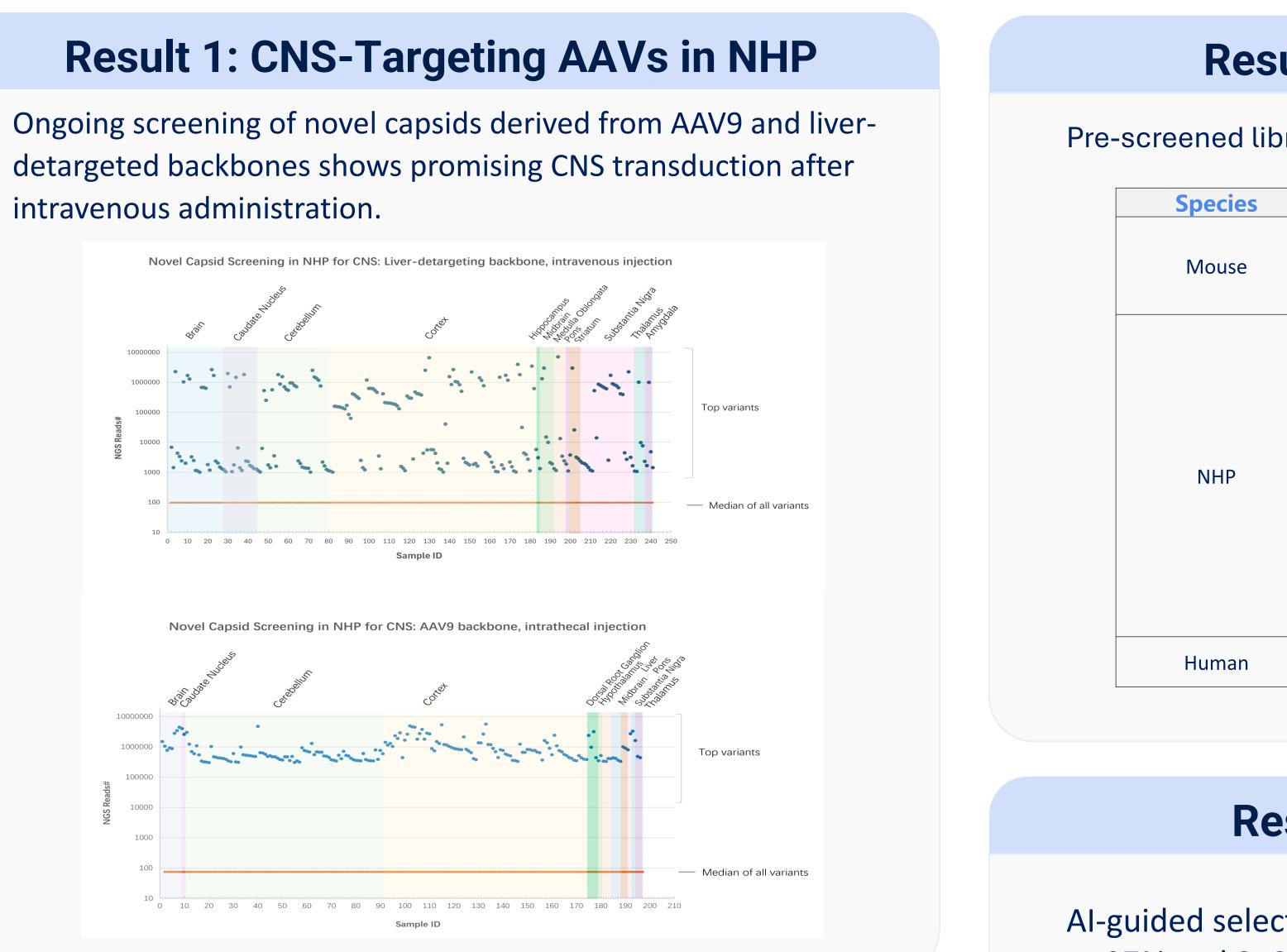


Li C, Samulski RJ. Engineering adeno-associated virus vectors for gene therapy. Nat Rev Genet. 2020 Apr;21(4):255-272. doi: 10.1038/s41576-019-0205-4. Epub 2020 Feb 10. PMID: 32042148. Li X, Wei X, Lin J, Ou L. A versatile toolkit for overcoming AAV immunity. Front Immunol. 2022 Sep 2;13:991832. doi:10.3389/fimmu.2022.991832. PMID: 36119036; PMCID: PMC9479010. Wu Z, Asokan A, Samulski RJ. Adeno-associated virus serotypes: vector toolkit for human gene therapy. Mol Ther. 2006 Sep;14(3):316-27. doi: 10.1016/j.ymthe.2006.05.009. Epub 2006 Jul 7. PMID: 16824801.

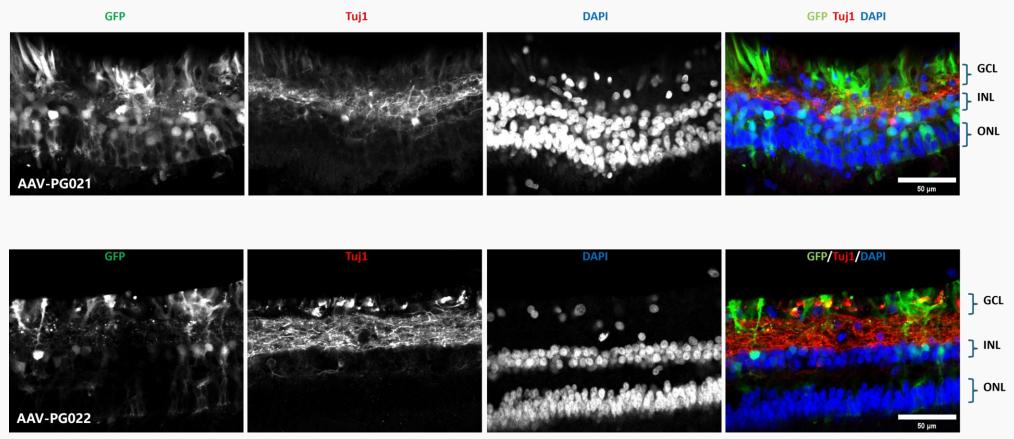
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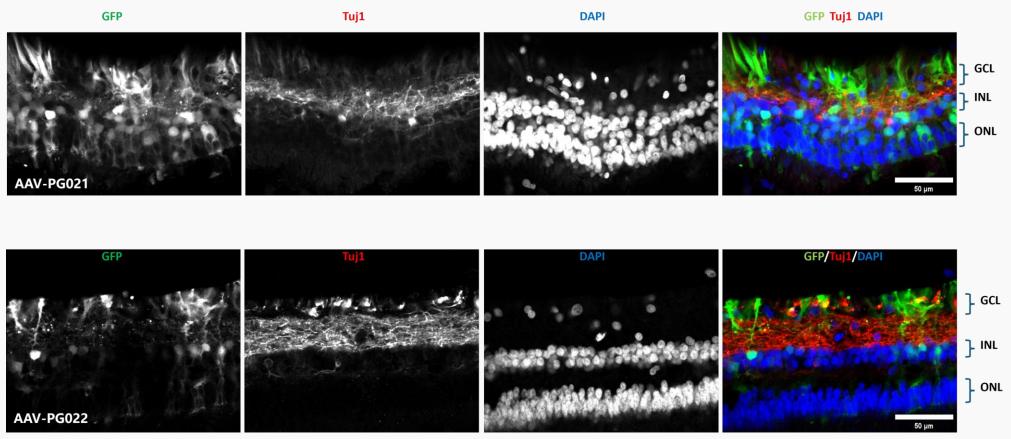
🛃 Ye Bu, Daniel Mchugh, Amos Gutnick, Xin Swanson, Irene Song, Paul Li* **O** PackGene Biotech, Houston, TX 77054 Corresponding author[#]: *irene.song@packgene.com*

Reference



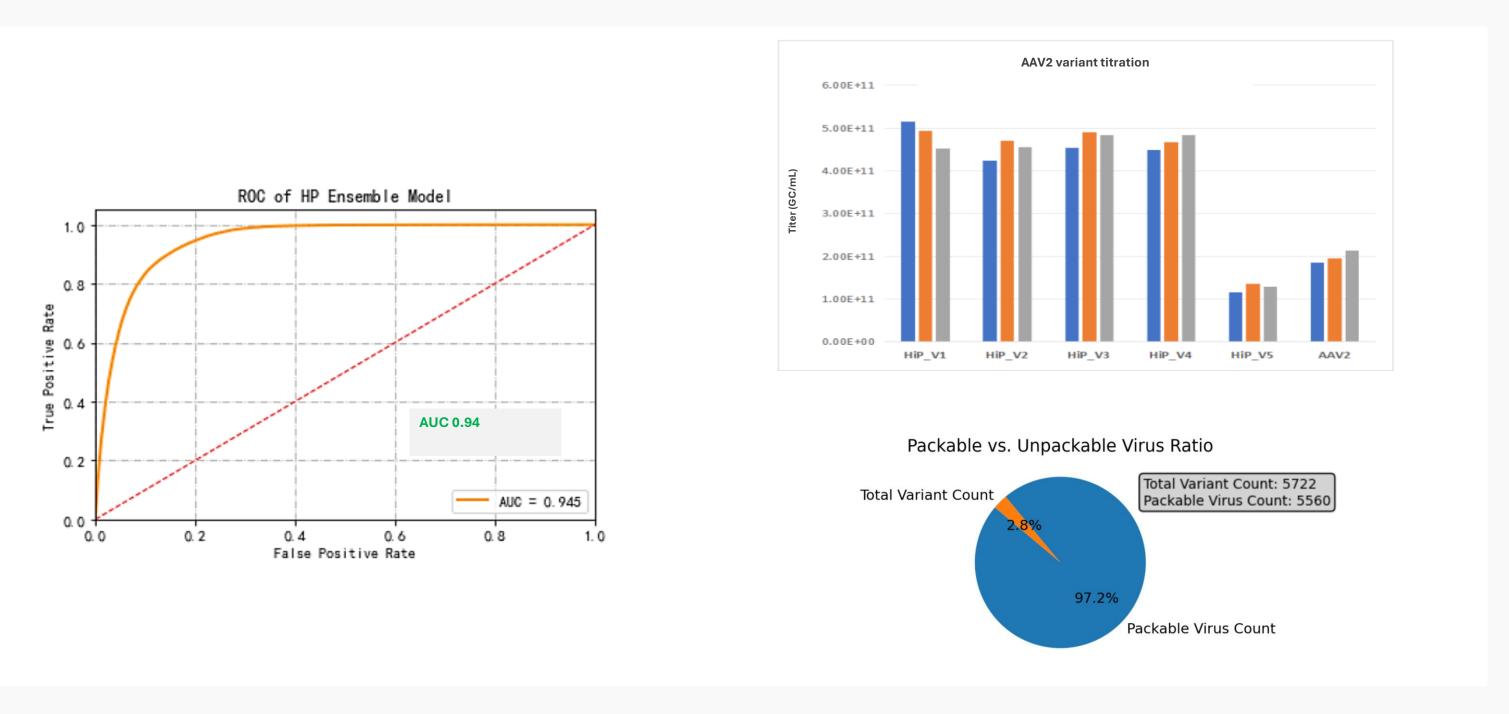
Variants PG021 and PG022 demonstrated effective transduction of retinal ganglion cells (RGCs), Müller glia, and INL cells.



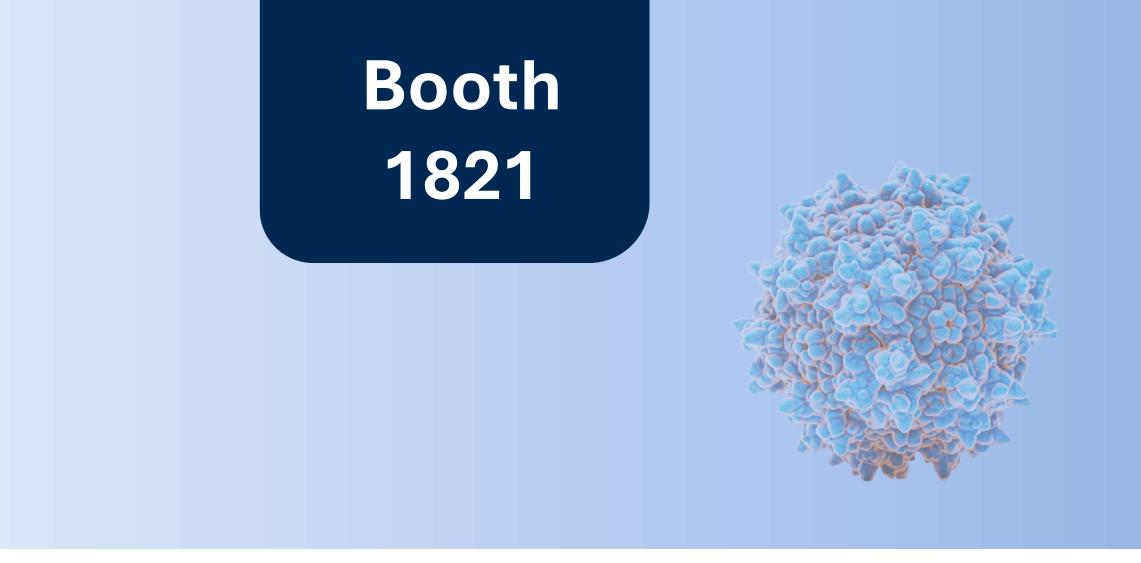


Result 2: Retinal-Targeting Capsids in NHP

PG021, PG022 - Retina targeting capsid screened from NHP



High-yield AAV2 variant with tissue specificity generated by AI model



Result 3: Broad Tissue-Specific Capsid Library

Pre-screened libraries yielded optimized capsids targeting CNS, muscle, and ocular tissues.

	Organ	Injection	Novel capsid
	CNS	Intravenous injection	PG008
	Muscle	Intravenous injection	PG007
	Muscle	Intravenous injection	PG007(screened from mouse but works well in NHP) PG0016,PG017, PG018, PG019, PG020
	CNS	Intrathecal injection	5 high potential variant in validation
		Intravenous injection	
	Eye/Cochlea	Local injection therapy	PG021, PG022, PG023
	Joint	Local injection therapy	2 nd round NGS data analysis
	Liver, Kidney, Lung	Intravenous injection	2 nd round screening
	T-cell	Infection	PG009, PG010, PG012, PG013, PG014

Result 4: Manufacturability Prediction Model

Al-guided selection produced high-yield AAV2 variants with packaging efficiencies up to 97% and 2–3x higher titers than wildtype.